

HOSTED BY



Contents lists available at ScienceDirect

Journal of King Saud University – Science

journal homepage: www.sciencedirect.com

Original article

Ganoderma multistipitatum sp. nov. from Chir pine tree (*Pinus roxburghii* Sarg.) in Pakistan



Shakil Ahmed^a, Muhammad Awais^b, Muhammad Muzzammal Sadiq^c, Aisha Umar^{a,*}, Laurent Dufossé^d, Muhammad Tajammal Khan^{e,*}, Jawaher Alkahtani^f, Rania M. Mahmoud^g

^a Plant Ecology and Environmental Biology Research lab, University of the Punjab, Lahore 54590, Pakistan

^b Avicenna Medical College, University of Health Sciences, Lahore, Pakistan

^c Rural Health Center Akhtar Abad, Tehsil Renala Khurd, District Okara, Pakistan

^d Université de La Réunion, Laboratoire CHEMBIOPRO (Chimie et Biotechnologie des Produits Naturels), ESIROI Département agroalimentaire, 15 avenue René Cassin, F-97490 Saint-Denis, France

^e Division of Science and Technology, Department of Botany, University of Education Lahore 5600, Pakistan

^f Department of Botany and Microbiology, College of Science, King Saud University, Riyadh 11451, Saudi Arabia

^g Department of Botany, Faculty of Science, University of Fayoum, Fayoum, Egypt

ARTICLE INFO

Article history:

Received 16 June 2022

Revised 28 November 2022

Accepted 5 January 2023

Available online 11 January 2023

Keywords:

Ganodermataceae

New species

ITS

Morphology

Phylogeny

Pinus roxburghii

ABSTRACT

Background: *Ganoderma* species are in state of flux in Pakistan as well as in the world. The diversity of this wood-inhabiting basidiomycete in Pakistan is still poorly known. A few species have been introduced from the country; even the country is rich with many *Ganoderma* species, which still need identification. **Methods:** *Ganoderma multistipitatum* sp. nov. is described from Lahore, Punjab Pakistan by using the morpho-anatomical (via naked eye and compound microscope) characters and molecular data from the ITS regions of DNA. The specimens used in this study were collected from the Botanical Garden of Government College University Lahore. The *G. multistipitatum* sp. nov. is, to date, only known from Lahore on Chir pine tree.

Results: Morphologically, this species is characterized by laccate orange, brown basidiomata, maroon, brown multi stipitate, and anatomically by ellipsoid basidiospores ($10.24\text{--}11.2 \times 5.3\text{--}5.4 \mu\text{m}$) and trimitic hyphal system. The *G. multistipitatum* sp. nov. is 21st species from the genus *Ganoderma* known from Pakistan. This new species is also described with photographs, line drawings and compared with nearby taxa of the phylogenetic tree constructed by software MEGA10. This species resembled with typical *G. lucidum*, but the ITS DNA sequences of this species were much far away from *G. multistipitatum* sp. nov. ones. **Conclusion:** The phylogeny based on ITS sequences supported this as a new distinct species, which finally compared with closely matrixed *G. multipileum* and related allies. The aim of this study was to introduce the new species from genus *Ganoderma*, which is a major contribution in biodiversity of the world.

© 2023 The Author(s). Published by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

1. Introduction

Ganoderma P. Karst. is a cosmopolitan genus and causes root and basal stem rot diseases (da Silva Coelho-Moreira et al., 2018;

Luangharn et al., 2019). *Ganoderma* is a basidiomycete genus, which belongs to the family Ganodermataceae (Richter et al., 2015; Tchotet Tchoumi et al., 2018). Chir pine (*Pinus roxburghii*) is widely planted for timber in its native area, being one of the most important trees. These trees are medicinal and found in Himalayan regions of Bhutan, Nepal, Kashmir, Sikkim, Tibet, and northern areas of Pakistan (Tripathi et al., 2008). This tree belongs to family Pinaceae commonly known as Chir Pine (Bissa et al., 2008). *Pinus roxburghii* has many ethnobotanical and medicinal uses (Kaushik et al., 2013).

Taxonomy of *Ganoderma* is polyphyletic based on pleomorphic morphology of basidiomata and geographical distribution (Wasser et al., 2006). Different phenetic plasticity of morpho-anatomical features hinder the correct identification of species (Hong et al.,

* Corresponding authors.

E-mail addresses: ash.dr88@gmail.com (A. Umar), tajammalkhan5@gmail.com (M. Tajammal Khan).

Peer review under responsibility of King Saud University.



Production and hosting by Elsevier

<https://doi.org/10.1016/j.jksus.2023.102554>

1018-3647/© 2023 The Author(s). Published by Elsevier B.V. on behalf of King Saud University.

This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

2004). Morphological and molecular approaches have been adopted for the *Ganoderma* species delineation and identification. Sequences of ITS rDNA have been commonly analysed. This analysis proved efficient in delimiting and resolving the species within *Ganoderma* species complexes (Mukhtar, 2019). The ITS fungal barcode is highly conserved intra-specifically, but acts as a variable for different species used in taxonomy (Fryssouli et al., 2020). The difference of ITS regions less than 2% nucleotides is considered valuable for intraspecific variation in genus *Ganoderma*, which facilitate the separation of certain species (Badotti et al., 2017).

Regarding the advancement in taxonomy of *Ganoderma* species, many new records and species are still being discovered and discovering from Asia (Umar et al., 2021a,b; Gafforov et al., 2020; Mukhtar, 2019). This genus is poorly studied in many regions of Asia like Pakistan, from which only *G. ahmadii* Steyaert, *G. lucidum* (Curtis) P. Karst., *G. applanatum* (Pers.) Pat., *G. flexipes* Pat., *G. resinaceum* Boud., *G. tornatum* (Pers.) Bres., *G. tsugae* Murrill, *G. australe* (Fr.) Pat., *G. boninense* Pat., *G. chalcone* (Cooke) Steyaert, *G. curtisii* (Berk.) Murrill, *G. lipsiense* (Batsch) G.F. Atk., *G. praelongum* Murrill, *G. multicornum* Ryvarden, *G. multiplicatum* (Mont.) Pat., and *T. colossus* (Fr.) Murrill [as *G. colossus* (Fr.) C.F. Baker] are known (Irshad et al., 2012; Steyaert, 1972; Ahmad, 1972; 1956) without morphological description and molecular data, but these estimations solely relied on morphological criteria of identification. Recently *G. leucocontextum* (Umar et al., 2021a), *G. multipileum* Ding Hou., *G. gibbosum* (Blume & T. Nees) Pat. (Umar et al., 2022), and *G. pakistanicum* A. Umar (Umar et al., 2021b) are published with morpho-anatomical and molecular data basis. The description of this new species brings a major contribution in the diversity of genus *Ganoderma* explored from Pakistan. Therefore, a convincing update is needed in taxonomy of *Ganoderma* species from Pakistan with the support of morphological and molecular data.

In the past, discrepancies were observed in taxonomy and classification of genus *Ganoderma*, so the objective of this study is the identification of new *Ganoderma* sp. by ITS marker, which facilitates the correct identification of the specimens. The morpho-anatomical studies and phylogenetic tree supported this work in species identification. Here we explored and introduced the 21st *Ganoderma* species, which highly contributed to the biodiversity of genus *Ganoderma*.

2. Materials & Methods

2.1. Studied materials

The specimens studied here were collected in 2019 by the fourth author from Botanical Garden of Government College University, Lahore, Punjab, Pakistan (31°49'81"N 73°30'44"E), elevation 217 m a.s.l. This garden is also called "Secretariat of Pakistan Botanic Gardens Network" covered with the major plant groups. Pteridophytes (*Nephrolepis*, *Pteris*, *Adiantum*), Gymnosperms (Cycads), *Pinus roxburghii*, *Ginkgo biloba*, *Cupressus*, and many other important tree plants of Angiosperms. The garden is also enriched with native trees of Punjab (Amin, 2013). This *Ganoderma* species grows on *Pinus roxburghii*. The average annual rainfall of Lahore is 607 mm and the temperature 24°C (Shirazi and Kazmi, 2016).

2.2. Morphological examination

Microscopic structures observed from cross sections of the basidiome that were soaked in KOH (5%), stained with Congo red (1%), and viewed under a MX4300H compound light microscope. Data of anatomical features were recorded at a magnification of 100X. Basidiospores were presented as length × width (Nagy et al., 2010) and measured without taking into account the apical

umbo when not shrunk. The morphological descriptions of the microscopic features were in part following Torres-Torres & Guzmán-Dávalos (Torres-Torres and Guzmán-Dávalos, 2012).

2.3. DNA extraction

Modified CTAB procedure was followed to extract total genomic DNA from dried specimens (Doyle and Doyle, 1987). The ITS1 + 5.8S + ITS2 rDNA region (henceforth referred to as the ITS region) was used to study the target specimens. This region was amplified by using primers ITS1 and ITS2 (White et al., 1990). Reaction mixtures (20 µl) contained 0.5 µl template DNA, 8.5 ml distilled water, 0.5 µl of each primer, and 10 ml PCR mix [DreamTaqGreen PCR Master Mix (2 X), Fermentas]. Amplification conditions were 35 cycles of 95°C for 30 s, 52°C for 30 s, and 72°C for 1 min, followed by a final extension at 72 °C for 10 min. Amplified PCR products were purified and sequenced by TSINGKE Co. Ltd. (China).

2.4. Data analysis by sequence alignment and molecular phylogeny

The ITS data set comprised DNA sequences of specimens from Pakistan and related species. The consensus of sequence was generated from both ITS1 and ITS2 in BioEdit vers. 7.2.5 (Hall, 1999) and then homology searches were performed at the NCBI web site using BLASTn. Additionally, ITS sequences of related species were downloaded from GenBank (<https://www.ncbi.nlm.nih.gov/genbank/>) and literature. All sequences were automatically aligned with MAFFT and manually adjusted using BioEdit (Tamura et al., 2011). The phylogenetic tree was constructed MEGA10 software to run the ML (maximum likelihood phylogenetic analysis) with 1000 bootstrap replicates (Tamura et al., 2011); branches with less than 50% bootstrap support were collapsed to matrix the species in which *Amaurodema rude* selected as an outgroup. The consensus of both sequences were finally deposited in GenBank for authenticity and reference.

3. Results

3.1. Phylogenetic inference

The ITS dataset enclosed 130 sequences of 47 taxa additionally two sequences of *Ganoderma multistipitatum* (GenBank No. ON032992, ON032991) and one from *Amaurodema rude* (outgroup), which was analyzed by maximum likelihood (ML) method. Our new species sequences have maximum query with undescribed *Ganoderma* sp., during the initial blast. But in phylogeny, these un-described *Ganoderma* species of initial blast were matrixed away from the sequences of this new species. So, these species were removed in the final phylogenetic tree. Cut the ambiguous ends of the ITS regions. Reference sequences from GenBank of closely related species were also combined in phylogeny. The multiple sequence alignment was performed by an online program MAFFT (Katoh and Standley, 2013). The resulting alignment was manually edited in BioEdit (Tamura et al., 2011), and the evolutionary model for nucleotides that fit best in data sets were selected.

Since the tree topologies resulting from the ML analyses is presented, along with the statistical bootstrap values (99%) (Fig. 1). This group formed a clade nearby *Ganoderma multipileum*, *G. martinicense*, and *G. parvulum*. In this study, novel species of *Ganoderma* found on *Pinus roxburghii* was described on the appearance of basidiome shape, pileus surface and color along with molecular phylogeny.

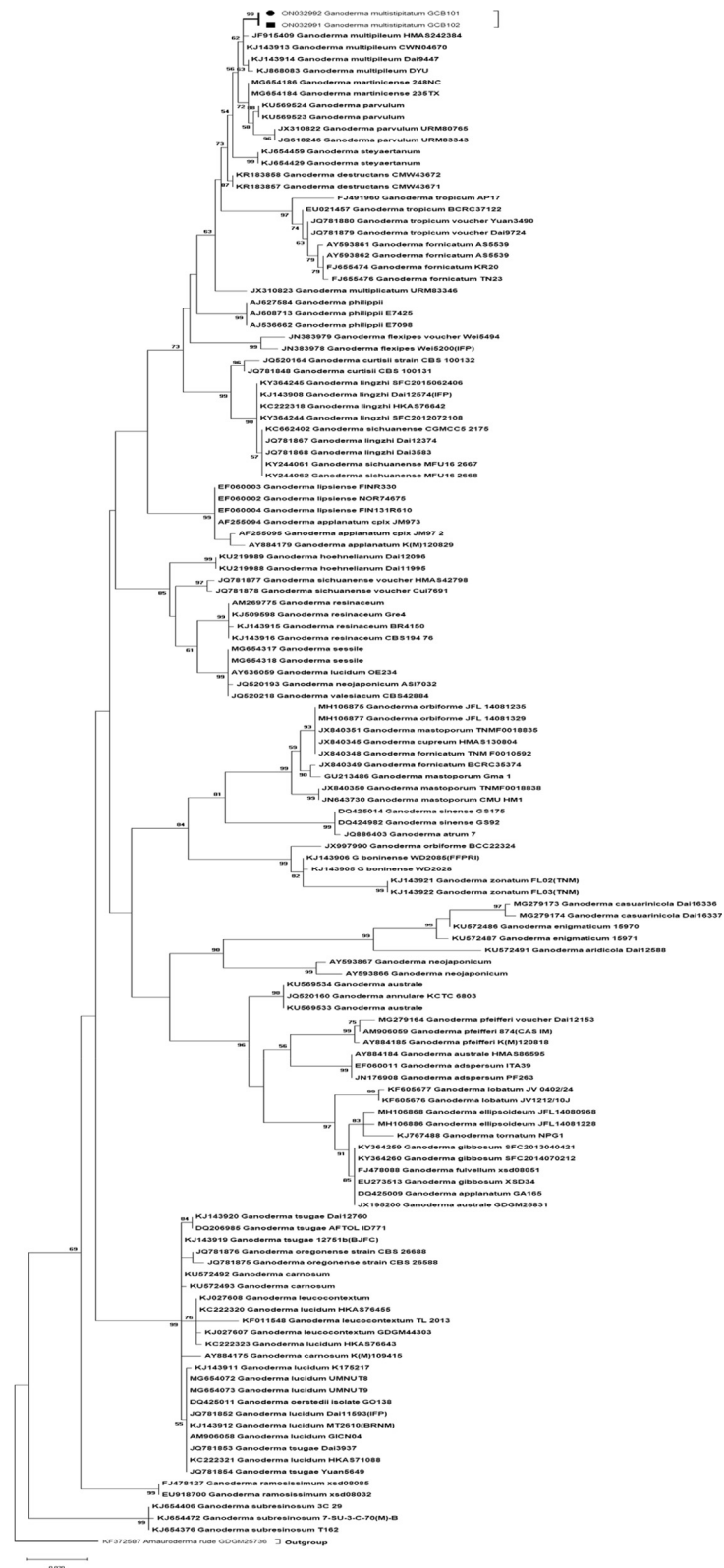


Fig. 1. Phylogenetic tree of *Ganoderma multistipitatum* (ITS rDNA sequences generated by maximum likelihood). Outgroup was *Amauroderma rude*. Bootstrap values (>50 %) are shown at the branches (New species represented by black dot and box).

3.2. Taxonomy of *Ganoderma multistipitatum* A. Umar, sp. nov.

3.2.1. Diagnosis:

Morphologically, *Ganoderma multistipitatum* sp. nov. resemble the species in subgenus *Ganoderma* P. Karst. It is characterized by annual, dimidiate and broadly orange brown laccate basidiome. The unique character of this species is the group of stout stipes give rise to one basidiome. Context was thick with white resinous

melanoid bands and white greyish growth zones. Trimitic hyphal system and basidiospores are ellipsoid, guttulated, bitunicate, brown, and with sub free minute inter-walled pillars (Figs. 2-4).

3.2.2. Holotypus

PAKISTAN. PUNJAB PROVINCE: Lahore, GC University, Lahore, Punjab, Pakistan, (31.4981° N 73.3044° E), elevation 217 m a.s.l,



Fig. 2. Macroscopic structures of *Ganoderma multistipitatum* (A–D GCB101). A–B. Basidiomata. C. Group of stipes. D. Lower view of basidiome. E. Pores surface. F. Section of context and tubes. Bars = 2 cm (A–D), 2 mm (E, F). (Photo by Aisha Umar).

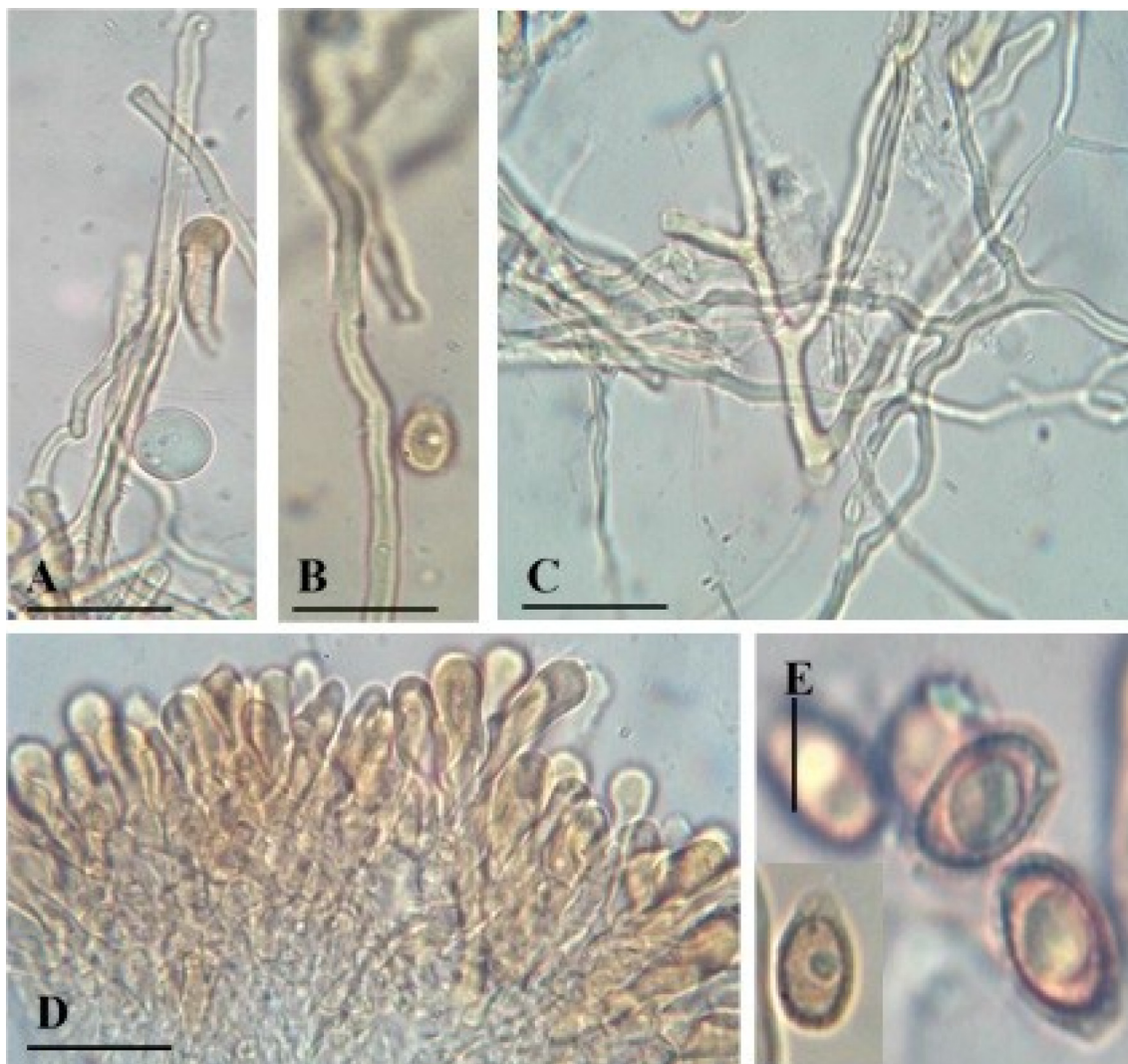


Fig. 3. Microscopic structures of *Ganoderma multistipitatum* (A–E GCB101). **A–C.** Hyphae (A. Generative), (B. Skeletal), (C. Binding). **D.** Cells of crustohymeniderm. **E.** Basidiospores. Bars = 5 µm (A–C), 10 µm (D–E).

attached to trunk of *Pinus roxburghii*, 15 July 2018, Aisha Umar, (holotype GCB101). GenBank: ITS = ON032992.

3.2.3. Etymology

The species epithet refers to “*multistipitatum*” represents the “group of stipes” give rise to one basidiome.

3.3. Morphological description

Basidiomata stipitate, verrucose, laccate cinnamon orange brown; **Stipe** 8.5–8.7 × 5.9–6.2, multistipitate, laccate, maroon brown to blackish brown, soft with hard crust; **Margin** 1.5–1.6 cm, broader, thick, prominent milky white, obtuse to incurved; **Pileus** 24–26 × 10–11 cm, 0.5–0.6 mm thickness, glossy, bumps, protuberances, reniform, dimidiate to flabelliform, radially wrinkled, soft flesh with hard crust; **Pores** 120–153 × 223–255 µm, white (fresh) to light brown (bruised), subcircular; “**Cutis**/palisado-derm 25.7–32.9 × 7.59–6.9 µm, thick-walled, clavate, palisade elements, yellowish brown”; **Tubes** 0.9–1.0 mm long, non-stratified, light brown; **Context** 1.4–1.6 cm long, light brown, dry, fibrous, soft, melanoid bands, white greyish growth zones; **Basidiospores** 10.24–11.2 × 5.3–5.4 µm, ellipsoid, bitunicate, exosporium hyaline, pigmented, thick, green, inner episporium, echinulae green

eusporium, guttulated, highly thick inter-walled pillars, apically truncate, tapering myxosporium end”; **Hyphal System** 1) generative hyphae (septate, clamped, colorless, thin-walled), 2) skeletal hyphae (thick-walled, colorless, unbranched or few branches with distal end), 3) binding hyphae (arboriform, brown, thick-walled, much-branched”).

3.4. Ecology and habitat

End of June to start of August, this species occurs on trunk of Chir tree forest of Punjab Province.

3.5. Additional specimens examined

PAKISTAN, PUNJAB PROVINCE: Lahore, (31.4981°N 73.3044°E), elevation 217 m a.s.l, attached to a tree trunk of *Pinus roxburghii*, 25 July 2019, Aisha Umar, (paratype GCB102). GenBank: ITS = ON032991.

4. Discussion

Ganoderma multistipitatum sp. nov. characterized by annual multistipitate, bumps (protuberances), dimidiate to flabelliform,

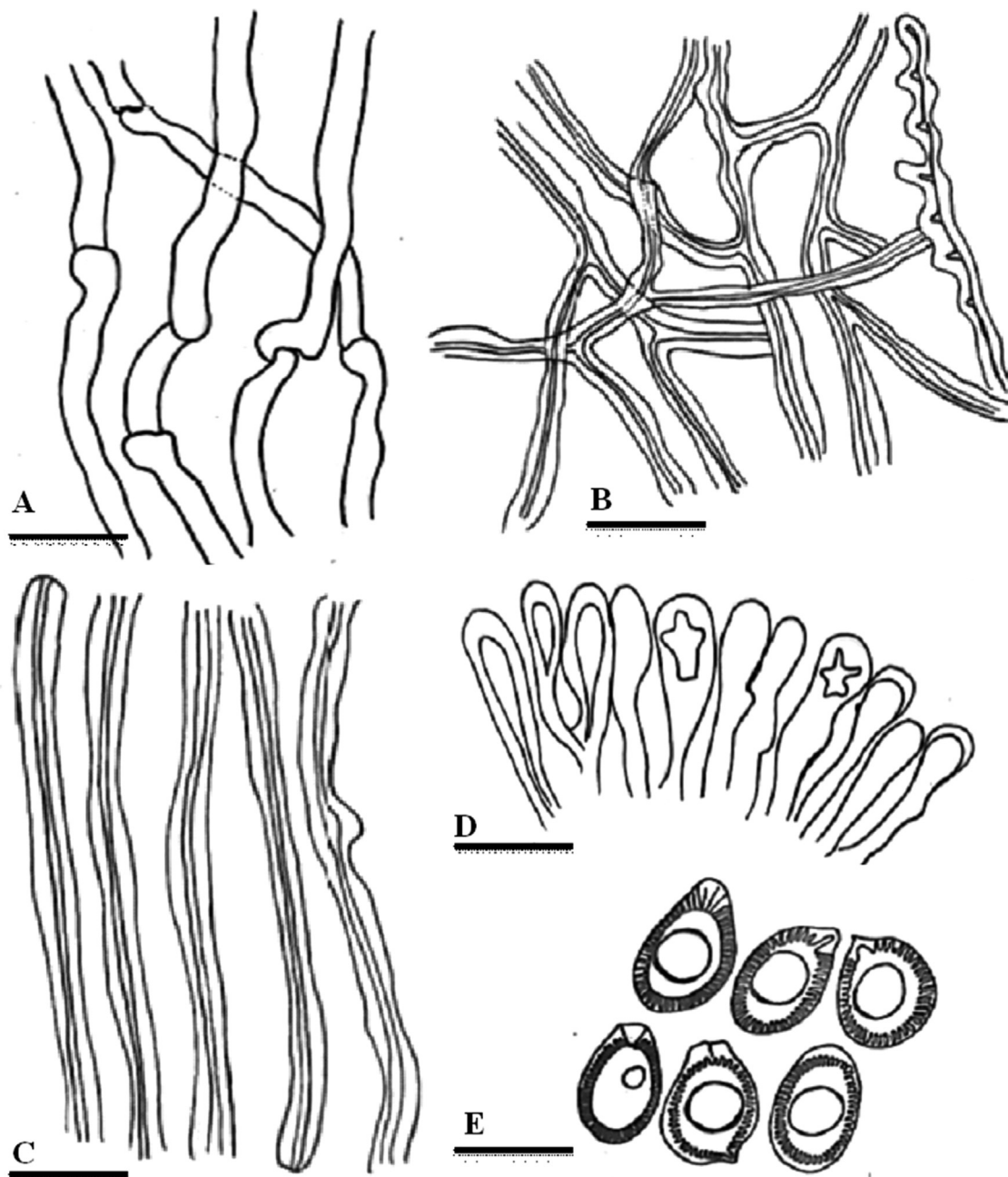


Fig. 4. Line drawings of anatomical characters of *Ganoderma multistipitatum*. A. Generative hyphae. B. Binding hyphae. C. Skeletal hyphae. D. Cells of crustohymeniderm. E. Basidiospores. Bars = 5 μ m (A–C), 10 μ m (D–E). (Drawn by Aisha Umar)

radially wrinkled, corky thick and shiny basidiome with very thick white margins. The light brown context with white melanoid band and trimitic hyphal system was also observed in this species.

This new species was closely related to lowland Taiwan and Chinese *G. multipileum* (Fig. 1). The main distinguishing features of *G. multipileum* also observed in Pakistani *G. multistipitatum* sp. nov. specimens are stalked basidiomata, thin crust, truncate ovoid basidiospores (8.2–9.4 $\times$ 4.7 μ m) (Wang et al., 2009), with minute inter-walled pillars, heterogeneous context (clay buff to fulvous) with melanoid bands, and concentric growth zones on basidiomata. Enlarged bulbous hyphal ends were observed by Cao et al. (2012) in *G. multipileum*, but not observed in the specimens of this study. Following Chang (1983), the features of stalked basidiomata and pilei growing together of *G. multipileum* are not

observed in *G. multistipitatum* sp. nov., whereas group of stipes are useful for the circumscription of this new specimens.

The other closest species of *G. multistipitatum* sp. nov. in phylogenetic tree of this study was *G. martinicense*, *G. parvulum* and *G. tropicum* (Fig. 1). *G. martinicense* is relative to *G. multipileum*, while geographically distant species only known from America (Loyd et al., 2018). *G. martinicense* has similarity with *G. multistipitatum* sp. nov. in melanoid bands and concentric growth zones in dark brown context (light brown in our specimens), dark red to black pseudostipe (group of stout stipes in our specimens), and longer coarse ornamented basidiospores (Welti and Courtecuisse, 2010). The *G. martinicense* has golden yellow pileal surface, whereas light sugar cane brown to brown or maroon, brown pileal surface found in the specimens of this study.

Ganoderma parvulum Murrill is morphologically very distinct from *G. multipileum* (Correia de Lima Júnior et al., 2014) and *G. multistipitatum* sp. nov. The *G. parvulum* is known from North and South America, possess light to pale context with thick black resinous bands contrary to Pakistani new species (Correia de Lima Júnior et al., 2014; Cabarro-Hernández et al., 2019). *G. parvulum* was synonym of *G. stipitatum* (Gottlieb and Wright, 1999), because both are same species (Ryvarden, 2004), but preference gives to *G. parvulum* on *G. stipitatum* (Murrill, as described earlier. The basidiospores size of Brazilian *G. parvulum* and *G. stipitatum* is similar but smaller ($8-10 \times 5-6 \mu\text{m}$) than our new species. Free to sub free and very thin pillars in endosporium are found in *G. parvulum* and *G. multistipitatum* sp. nov. (Cabarro-Hernández et al., 2019). The black thick resinose band is present in context of *G. parvulum*, while white band present in this new species. The clavate shape of cuticular cells are similar in both *G. parvulum* and *G. multistipitatum* sp. nov.

Ganoderma tropicum from tropical and subtropical areas of Asia is dissimilar to *G. multistipitatum* sp. nov. in morphology, because this species has concentric growth zones on the pileus, while absent in this species (Zhou et al., 2015). Both species has melanoid bands in the contextum. The contextum is homogeneous and fulvous in *G. tropicum* and *G. multistipitatum* sp. nov. The cuticular cells are mostly clavate, irregular, often with blunt protuberances or outgrowths in *G. tropicum*, whereas no growth found in *G. multistipitatum* sp. nov. The basidiospores are coarsely echinulate, ellipsoid to broadly ellipsoid and smaller ($8.8-10.7 \times 5.5-6.3 \mu\text{m}$) than our new *Ganoderma* species (Zhou et al., 2015).

Careful observations on morpho-anatomy differentiated the *Ganoderma multistipitatum* sp. nov. from *G. multipileum*, *G. tropicum*, *G. parvulum*, and *G. martinicense*. Furthermore, these are all phylogenetically distant from the specimens of this study.

5. Conclusion

Ganoderma multistipitatum sp. nov. represents the 21st *Ganoderma* species (accession numbers ON032992, ON032991) described from Pakistan. The wood-inhabiting basidiomycete diversity is still poorly known in Pakistan. A systematic survey of polypores in Pakistan is strongly needed. Taken the special geographic position of this region into consideration, this kind of survey improves the knowledge about wood-inhabiting polypore diversity of the world.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgment

We would like to pay a special thanks to the director of Institute of Botany, University of the Punjab, Lahore Pakistan, who allowed to do this work and facilitated in many ways. The authors extend their appreciation to the Researchers Supporting Project number (RSP2023R193), King Saud University, Riyadh, Saudi Arabia.

References

- Ahmad, S., 1956. Fungi of Pakistan, mon. 1. Lahore, Biological Laboratories, Government College.
 Ahmad, S., 1972. Basidiomycetes of west Pakistan, mon. 6. Lahore, Biological Laboratories, Government College.
 Amin, U.K., 2013. Guide to botanic garden G.C. University Lahore.

- Badotti, F., de Oliveira, F.S., Garcia, C.F., Vaz, A.B.M., Fonseca, P.L.C., Nahum, L.A., Oliveira, G., Góes-Neto, A., 2017. Effectiveness of ITS and sub-regions as DNA barcode markers for the identification of Basidiomycota (Fungi). *BMC Microbiol.* 17 (1), 1–2.
 Bissa, S., Bohra, A., Bohra, A., 2008. Antibacterial potential of three naked-seeded (Gymnosperm) plants. *Nat. Prod. Res.* 7 (5), 420–425.
 Cabarro-Hernández, M., Villalobos-Arámbula, A.R., Torres-Torres, M.G., Decock, C., Guzmán-Dávalos, L., 2019. The *Ganoderma weberianum-resinaceum* lineage: multilocus phylogenetic analysis and morphology confirm *G. mexicanum* and *G. parvulum* in the Neotropics. *Myckeys.* 59, 95–131 <http://dx.doi.org/10.3897/mycokeys.59.33182>.
 Cao, Y., Wu, S.H., Dai, Y.C., 2012. Species clarification of the prize medicinal *Ganoderma* mushroom “Lingzhi”. *Fungal Divers.* 56, 49–62. <https://doi.org/10.1007/s13225-012-0178-5>.
 Chang, T.T., 1983. Studies on biology of several species of *Ganoderma* in Taiwan [Master's thesis]. National Taiwan University, Taipei, p. 130.
 Correia de Lima Júnior, N., Baptista Gibertoni, T., Malosso, E., 2014. Delimitation of some neotropical laccate *Ganoderma* (*Ganodermataceae*): molecular phylogeny and morphology. *Rev. de Biol. Trop.* 62, 1197–1208.
 da Silva Coelho-Moreira, J., Brugnari, T., Sá-Nakanishi, A.B., Castoldi, R., De Souza, C. G., Bracht, A., Peralta, R.M., 2018. Evaluation of diuron tolerance and biotransformation by the white-rot fungus *Ganoderma lucidum*. *Fungal boil.* 122 (6), 471–478.
 Doyle, J.J., Doyle, J.L., 1987. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochem. Bull.* 19, 11–15.
 Fryssoulis, V., Zervakis, G.I., Polemis, E., Typas, M.A., 2020. A global meta-analysis of ITS rDNA sequences from material belonging to the genus *Ganoderma* (Basidiomycota, Polyporales) including new data from selected taxa. *Myckeys.* 75, 1–143. <https://doi.org/10.3897/mycokeys.75.59872>.
 Gafforov, Y., Ordynets, A., Langer, E., Yarasheva, M., de Mello Gugliotta, A., Schigel, D., Pecoraro, L., Zhou, Y., Cai, L., Zhou, L.W., 2020. Species diversity with comprehensive annotations of wood-inhabiting poroid and corticioid fungi in Uzbekistan. *Front. Microbiol.* 3047.
 Gottlieb, A.M., Wright, J.E., 1999. Taxonomy of *Ganoderma* from southern South America: subgenus *Ganoderma*. *Mycol. Res.* 103, 661–673. <https://dx.doi.org/10.1017/S0953756298007941>.
 Hall, T.A., 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symp. Ser.* 41, 95–98.
 Hong, S.G., Jung, H.S., 2004. Phylogenetic analysis of *Ganoderma* based on nearly complete mitochondrial small-subunit ribosomal DNA sequences. *Mycologia.* 96 (4), 742–755.
 Irshad, M., Anwar, Z., Gulfranz, M., Butt, H.I., Ejaz, A., Nawaz, H., 2012. Purification and characterization of α -amylase from *Ganoderma tsuagii* growing in waste bread medium. *African J. Biotech.* 11, 8288–8294. <https://doi.org/10.5897/AJB113643>.
 Katoh, K., Standley, D.M., 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol. Biol. Evol.* 30, 772–780.
 Kaushik, P., Kaushik, D., Khokra, S.L., 2013. Ethnobotany and phytopharmacology of *Pinus roxburghii* Sargent: a plant review. *J. Integr. Med.* 11 (6), 371–376.
 Loyd, A.L., Barnes, C.W., Held, B.W., Schink, M.J., Smith, M.E., Smith, J.A., Blanchette, R.A., 2018. Elucidating “*lucidum*”: Distinguishing the diverse laccate *Ganoderma* species of the United States. *PLoS One.* 13 (7), e0199738.
 Luangharn, T., Karunaratna, S.C., Mortimer, P.E., Hyde, K.D., Xu, J., 2019. Additions to the knowledge of *Ganoderma* in Thailand: *Ganoderma casuarinicola*, a new record; and *Ganoderma thailandicum* sp. nov. *Myckeys.* 59, 47.
 Mukhtar, T., 2019. Morphological characterization of *Ganoderma* species from Murree Hills of Pakistan. *Plant Prot.* 3 (2), 73–84 <https://doi.org/10.33804/pp.003.02.0128>.
 Nagy, G.L., Vágvolgyi, C., Papp, T., 2010. Type studies and nomenclatural revisions in *Parasola* (Psathyrellaceae) and related taxa. *Mycotaxon* 112, 103–141 <http://dx.doi.org/10.5248/112.103>.
 Richter, C., Wittstein, K., Kirk, P.M., Stadler, M., 2015. An assessment of the taxonomy and chemotaxonomy of *Ganoderma*. *Fungal Divers.* 71 (1), 1–15.
 Ryvarden, L., 2004. Neotropical polypores: Part 1: Introduction, *Ganodermataceae* & *Hymenochaetaceae*. *Fungiflora*.
 Shirazi, S.A., Kazmi, J.H., 2016. Analysis of socio-environmental impacts of the loss of urban trees and vegetation in Lahore, Pakistan: a review of public perception. *Ecol. Process.* 5 (1), 1–2.
 Steyaert, R.L., 1972. Species of *Ganoderma* and related genera mainly of the Bogor and Leiden Herbaria. *Persoonia.* 7, 55–118.
 Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., Kumar, S., 2011. MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol. Biol. Evol.* 28, 2731–2739. <https://doi.org/10.1093/molbev/msr121>.
 Tchotet Tchoumi, J.M., Coetzee, M.P.A., Rajchenberg, M., Wingfield, M.J., Roux, J., 2018. Three *Ganoderma* species, including *Ganoderma dunense* sp. nov., associated with dying *Acacia cyclops* trees in South Africa. *Australas. Plant Pathol.* 47, 431–447. <https://doi.org/10.1007/s13313-018-0575-7>.
 Torres-Torres, M.G., Guzmán-Dávalos, L., 2012. The morphology of *Ganoderma* species with a laccate surface. *Mycotaxon.* 119, 201–216. <https://doi.org/10.5248/119.201>.
 Tripathi, P., Tewari, L.M., Tewari, A., Kumar, S., Pangtey, Y.P.S., Tewari, G., 2008. Gymnosperms of Nainital. Kumaun University, Nainital.
 Umar, A., Ahmed, S., Bashir, H., 2021a. *Ganoderma leucocontextum*, a new record from Pakistan. *Mycotaxon.* 136 (2), 529–539.

- Umar, A., Ahmed, S., Gafforov, Y., 2021b. *Ganoderma pakistanicum* sp. nov. (Ganodermataceae, Basidiomycota) from Pakistan. *Nova Hedwigia*. 113 (3–40), 531–543. https://doi.org/10.1127/nova_hedwigia/2021/0656.
- Umar, A., Ahmed, S., Guzmán-Dávalos, L., Cabarroí-Hernández, M., 2022. *Ganoderma multipileum* and *Tomophagus cattienensis*—new records from Pakistan. *Mycotaxon*. 137 (1), 135–151. <https://doi.org/10.5248/137.135>.
- Wang, D.M., Wu, S.H., Su, C.H., Peng, J.T., Shih, Y.H., Chen, L.C., 2009. *Ganoderma multipileum*, the correct name for ‘*G. lucidum*’ in tropical Asia. *Bot Stud* 50, 451–458.
- Wasser, S.P., Zmitrovich, I.V., Didukh, M.Y., Spirin, W.A., Malysheva, V.F., 2006. Morphological traits of *Ganoderma lucidum* complex highlighting *G. tsugae* var. *janniae*: the current generalization. *ARA Gantner Verlag, Ruggell*, pp. 1–187.
- Walti, S., Courtecuisse, R., 2010. The *Ganodermataceae* in the French West Indies (Guadeloupe and Martinique). *Fungal Divers.* 43, 103–126. <https://doi.org/10.1007/s13225-010-0036-2>.
- White, T.J., Bruns, T., Lee, S.J.W.T., Taylor, J., 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenies. *PCR protocols, a guide to methods and applications*. 18 (1), 315–322.
- Zhou, L.W., Cao, Y., Wu, S.H., Vlasák, J., Li, D.W., Li, M.J., Dai, Y.C., 2015. Global diversity of the *Ganoderma lucidum* complex (Ganodermataceae, Polyporales) inferred from morphology and multilocus phylogeny. *Phytochemistry*. 114, 7–15.